

APPLICATIONS

Reproducible LC-MS/MS Separation of 38 Amino Acids in HILIC Mode using a bioZen™ Glycan LC Column

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Introduction

Amino acids are the building blocks of proteins, making them an extremely well researched compound class. With thousands of naturally occurring amino acids available, separating and quantitating a mixture of amino acids can be challenging. While HPLC is a well characterized method for amino acid analysis, LC-MS analysis of amino acids poses several challenges: 1. Some compounds are very polar and are hard to retain by reversed phase HPLC, 2. Isomers can coelute and are difficult to differentiate from each other, and 3. While effective, HILIC based separation sometimes results in poor reproducibility. These challenges were overcome by combining the separation power of the bioZen Glycan HILIC phase LC column with SWATH® Acquisition, resulting in highly reproducible resolution and accurate quantitation and confirmation of 38 amino acids.

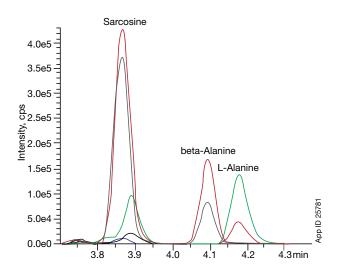
Materials and Methods

A sample of 38 amino acids was prepared using a standard stock mix (A9906, Sigma-Aldrich®), and used as 10x diluted for various experiments.

Buffers were prepared as follows:

Make 1 L of 100 mM Ammonium formate, then adjust pH to 3.1 with formic acid. Then, combine 900 mL of acetonitrile or water with 100 mL of the prepared buffer. Final buffer concentration should equal 10 mM in each bottle.

Figure 1.Baseline separation of isomers (SWATH MS/MS spectra)



HPLC Conditions

Column: bioZen Glycan, 2.6 μm
Dimensions: 100 x 2.1 mm
Part No.: 00D-4773-AN
Elution Type: Gradient

Mobile Phase: A: 10 mM Ammonium formate in Acetonitrile

B: 10 mM Ammonium formate in Water

Gradient: Time (min) % B 0.01 0 5 7 50 8 50 8.1 0 12 0 0

Flow Rate: 500 µL/min Temperature: 40 °C

System: TripleT0F® 6600 coupled with an ExionLC™ system (SCIEX)

MS Acquisition: SWATH® Acquisition

MS Detection: MS/MS Sample: 1. Urea

MS/MS

1. Urea

2. L-Creatine

3. 2-Aminoethanol

4. L-Tryptophan

5. L-Phenylalanine

6. L-Leucine

7. L-Isoleucine

8. Taurine

9. L-Methionine

10. L-Tyrosine

11. L-Proline

12. L-Valine

13. Gamma-aminobutyric acid

14. L-alpha-aminoisobutyric acid

15. L- Sarcosine

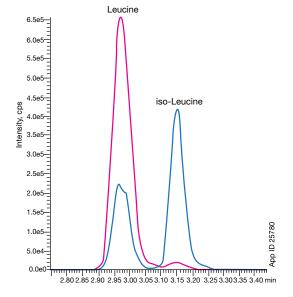
16. DI -3-aminoisobutyric acid

17. L- Sarcosine

15. L- Sarcosine
16. DL-3-aminoisobutyric acid
17. L-Arginine
18. 1-Methyl-L-histidine
19. Hydroxy-L-proline

20. 3-Methyl-L-histidine 21. L-Lysine 22. Beta-Alanine 23. L-Ornithine 24 Glycine 25. L-Threonine 26. Anserine 27. L-Histidine 28. L-Alanine 29. Hydroxylysine 30 L-Carnosine 31. L-Serine 32. L-Citrulline 33. L-Homocystine 34. L-Cystathionine 35. L-Cystine 36. L-Aspartic acid 37. 2-Aminoadipic acid

38. L-Glutamic acid





Results and Discussion

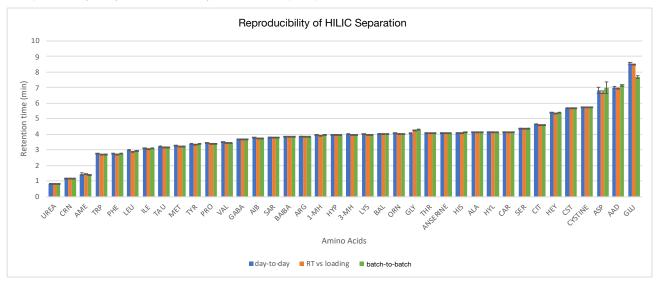
Due to the polar and hydrophilic nature of amino acids, HILIC chromatography was utilized to separate the 38 amino acid mixture. HILIC mode provided better retention and baseline isomeric separation as compared to the reversed phase separation of these compounds (**Figure 1**). Although HILIC mode was successful in separating the 38 amino acids in our analysis, it was imperative that the reproducibility of the separation be challenged. Inter-day, intra-day, lot-to-lot, and multiple injection volumes were studied to verify the robustness of our method (**Figure 2**). This was performed by measuring any fluctuation in run times under various conditions. The results of these studies are depicted in **Figure 2**, where the following conditions were studied:

- 1. 3 batches and 15 injections per batch
- Day-to-day evaluation which includes 4 days and 5 injections per day
- 3. Run time vs. loading evaluation (1, 2, 3, 4, and 5 μL injection volumes at 3 injections per volume)

Conclusion

Utilizing a highly reproducible bioZen Glycan LC column (HILIC phase) and a robust LC-MS/MS method, 38 amino acids were successfully separated. This methodology was tested for robustness under various conditions including inter- and intra-day, various media lots, and several different injection sizes to ensure that the separation was not only successful but also robust and reproducible. Furthermore, this methodology using MS detection, especially SWATH® Acquisition, allows for accurate amino acid quantitation in various biological matrixes

Figure 2. Reproducibility study of the bioZen™ Glycan LC column (HILIC)



Ordering Information

bioZen Columns (mm)				Biocompatible Guard Cartridges	
Phase	50 x 2.1	100 x 2.1	150 x 2.1	for 2.1 mm	Holder
bioZen 2.6 µm Glycan	00B-4773-AN	00D-4773-AN	00F-4773-AN	<u>AJ0-9800</u>	<u>AJ0-9000</u>





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