

APPLICATIONS

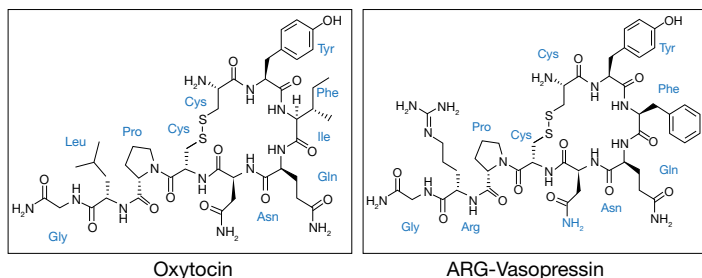
LC-MS/MS Analysis Oxytocin (OT) and ARG-Vasopressin (AVP) in Human Plasma/Serum using Strata[®]-X-CW Solid Phase Extraction (SPE) and a Luna[®] PFP(2) HPLC Column

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Introduction

Immunoassay, nano-LC systems, and 2D-LC-MS/MS methods are commonly used to reach such low detection levels for Oxytocin (OT) and Arginine-vasopressin (AVP); however, these methodologies also have limitations such as assay reproducibility issues, facility instrument limitations, and high method development and validation costs.

In this study, we provide an accurate and reproducible method to analyze both OT and AVP in one injection.



Materials and Methods

Reagents and Chemicals

Oxytocin, vasopressin, and octreotide standards were purchased from Sigma-Aldrich.

Experimental Conditions

Solid Phase Extraction Method Development

300 μ L of plasma/serum was treated with 300 μ L 4% phosphoric acid and 50 μ L of internal standard (500 ng/mL of Octreotide) in water and mixed. Strata-X, Strata-X-C, and Strata-X-CW were evaluated to determine the optimal solid phase extraction sorbent (**Figure 1**). For the final selected sorbent, Strata-X-CW, wash and elution solvents were optimized (**Figures 2 and 3**). Although Strata-X performed well for AVP, only Strata-X-CW was able to provide acceptable recoveries for both OT and AVP.

LC-MS/MS Conditions

LC-MS/MS was performed using a Luna 3 μ m PFP(2) 50 x 2.0mm HPLC column (00B-4447-B0) on an Agilent[®] 1200 LC system (Agilent Technologies, Palo Alto, CA, USA) with an upper pressure limit of 400 bar, equipped with a binary pump, autosampler and interfaced with an API 5000[™] triple quadrupole mass spectrometer (AB SCIEX, Framingham, MA, USA). The ionization source was electrospray ionization (ESI) analyzed in positive ion mode. Ionization source parameters and LC running conditions are indicated.

Figure 1. Response of OT and AVP using Various SPE Sorbents
SPE Sorbent Selection

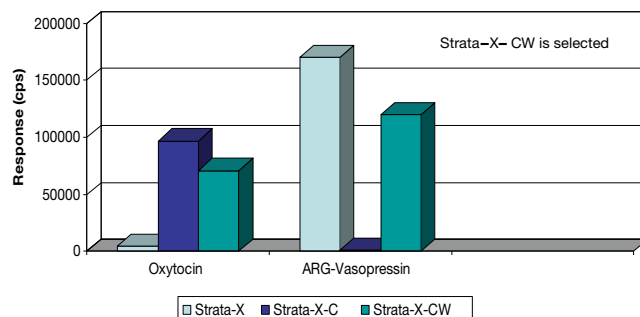


Figure 2. Response of OT and AVP of 2nd Wash Optimization using Strata-X-CW
2nd Wash Optimization

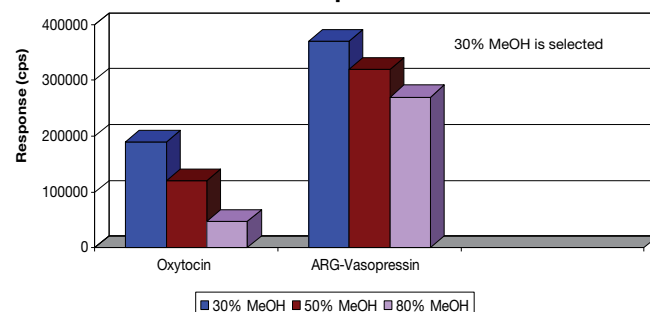
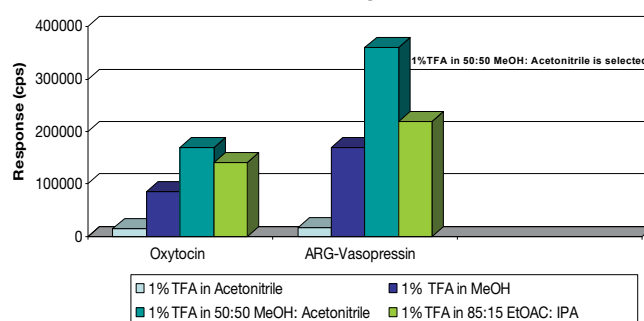


Figure 3. Response of OT and AVP Under Various Elution Strengths using Strata-X-CW
Elution Solvent Optimization



Optimized SPE Method

96-Well Plate: Strata-X-CW, 30 mg/well
Part No.: 8E-S035-TGB
Condition: 1 mL Methanol
Equilibrate: 1 mL Water
Load: Pretreated plasma/serum
Wash 1: 1 mL 4% H₃PO₄
Wash 2: 1 mL Methanol/Water (30:70)
Dry: 3 to 4 minutes under 10" Hg vacuum
Elute: 2x 0.75 mL 1% TFA in Acetonitrile/Methanol (50:50)
Dry down: Dry down completely under a stream of nitrogen @ 50 °C
Reconstitute: 100 μ L of 1% TFA in Mobile Phase A/Mobile Phase B (80:20)
Mobile Phase A: 0.1% Formic acid; B: Methanol/Acetonitrile (50:50) with 0.1% Formic acid

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Results and Discussion

Figure 4 and **Figure 5** show extracted ion chromatograms at QCL levels of 0.500 ng/mL. **Figure 6** shows a representative chromatogram at analyte levels of 500 ng/mL, showing that the Luna PFP(2) HPLC column provided good selectivity and sensitivity for both Oxytocin and ARG-Vasopressin in a single injection.

Table 2 shows the accuracy and precision of three QC levels. The results showed that the assay was accurate, precise, and reproducible. Linearity was determined to be acceptable from 0.2 - 500 ng/mL (**Figures 7** and **8**).

Ionization Source Parameters:

Gas 1 & Gas 2: 25
CAD: 8
Cur: 25
IS: 5500
Temp: 450
ihe: ON
DP: 120
EP: 10

Column: Luna® 3 µm PFP(2)

Dimensions: 50 x 2.0 mm

Part No.: 00B-4447-B0

Mobile Phase: A: 0.1 % Formic acid

B: Methanol/Acetonitrile (50:50) with 0.1 % Formic acid

Gradient: Time (min) B (%)

0 10
0.2 10
0.5 30
3.5 30
3.51 70
4 70
4.01 10
5 10

Flow Rate: 0.45 mL/min

Injection: 10 µL

Temperature: 40 °C

Detection: SCIEX® API 5000™

Table 1. MRM Transitions for OT and AVP

Analyte Peak Name	Q1 (Da)	Q3 (Da)	Dwell (msec)	CE	CXP
Oxytocin 1	1007.9	723.4	75	39	15
Oxytocin 2	504.8	86.2	75	35	15
Vasopressin 1	542.5	328.6	75	28	15
Vasopressin 2	542.5	120.1	75	28	15
Octreotide (IS)	510.5	120.1	75	30	13

Q1. Quantitation mass transitions

Q2. Confirmation mass transitions

Conclusion

In this study, we combined OT and AVP and analyzed them within a single method. To maximize assay recoveries, the 2nd strong wash and elution solvents have been optimized. We also provided stable MRM transitions for both OT and AVP, the assay dynamic range is 0.2 - 500 ng/mL for both compounds with precision less than 13.9 % and accuracy less than 7.4 %. This method would provide a good starting point for reference labs as an alternative approach to immunoassay. Although our resulting method was accurate at concentration ranges of 0.2 - 500 ng/mL, the LLOQ level could be decreased significantly if coupled with a nano-LC system or HRMS.

Figure 4. Oxytocin Extracted Ion Chromatograms, QCL (0.500 ng/mL)

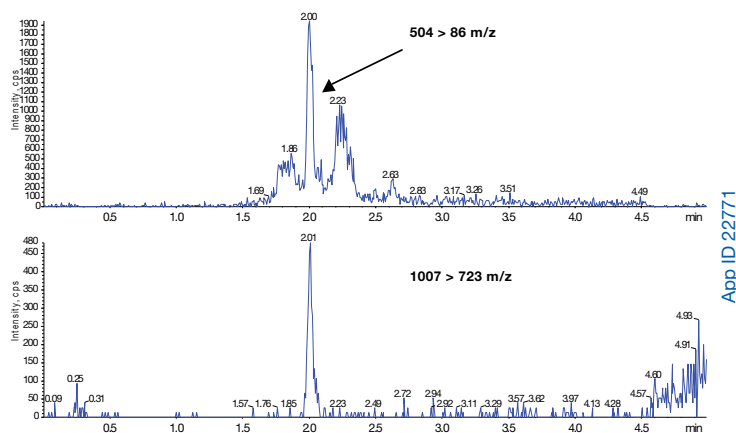


Figure 5. ARG-Vasopressin Extracted Ion Chromatograms, QCL (0.500 ng/mL)

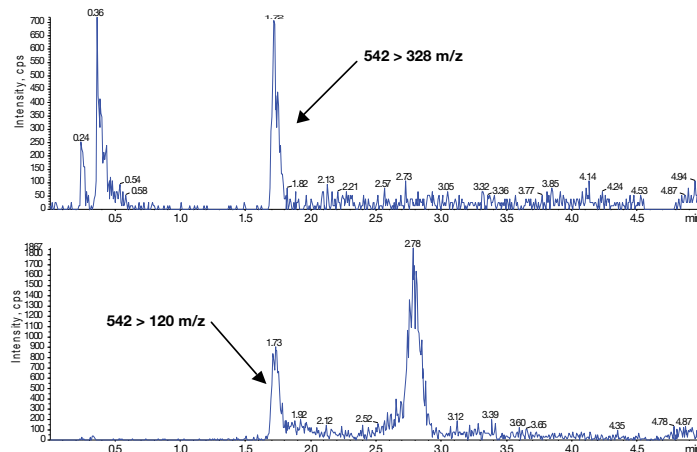
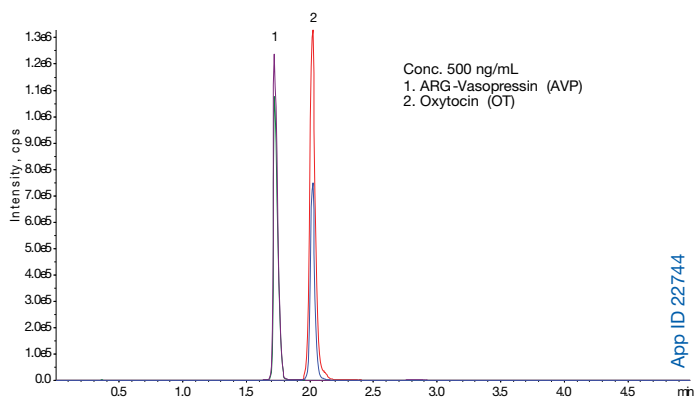


Figure 6. Representative Chromatogram of ARG-Vasopressin and Oxytocin



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Figure 7. Representative Curve of Oxytocin

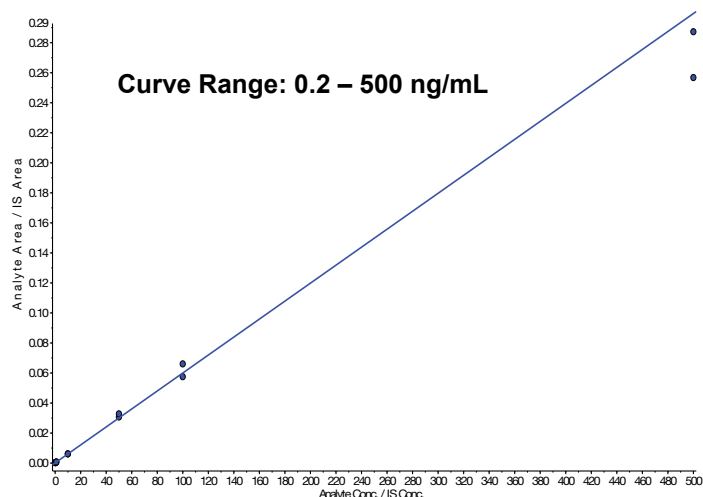


Figure 8. Representative Curve of ARG-Vasopressin

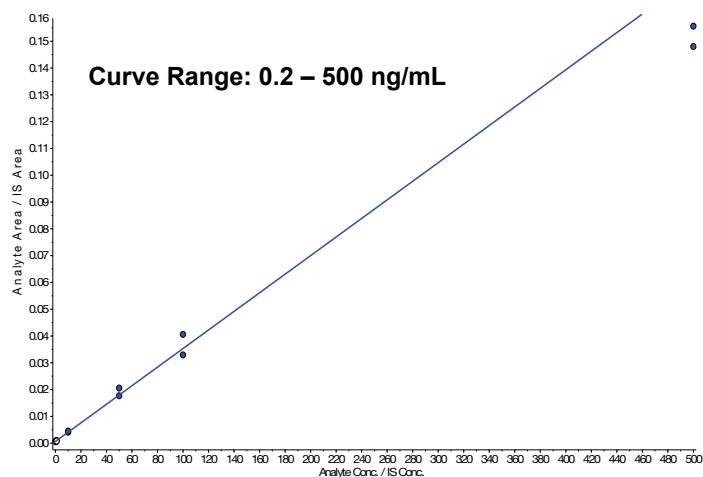


Table 2. Accuracy and Precision

	QCL	QCM	QCH
Nominal Conc. (ng/mL)	0.5	10	400
Oxytocin (OT)			
Mean Conc. Fund (ng/mL)	0.521	10.1	382
STDV	0.025	0.597	25.8
CV %	4.72	5.94	6.76
Accuracy (%)	104	101	95.5
ARG-Vasopressin (AVP)			
Mean Conc. Fund (ng/mL)	0.473	10.2	370
STDV	0.666	0.367	23.3
CV %	13.9	3.59	6.29
Accuracy (%)	94.5	102	92.6

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Strata®-X-CW SPE Ordering Information

Format	Sorbent Mass	Part Number	Unit
Tube			
	30 mg	8B-S035-TAK**	1 mL (100/box)
	30 mg	8B-S035-TBJ	3 mL (50/box)
	60 mg	8B-S035-UBJ**	3 mL (50/box)
	100 mg	8B-S035-FCH	6 mL (30/box)
	200 mg	8B-S035-FBJ	3 mL (50/box)
	200 mg	8B-S035-FCH	6 mL (30/box)
	500 mg	8B-S035-HBJ	3 mL (50/box)
	500 mg	8B-S035-HCH	6 mL (30/box)
Giga™ Tube			
	1 g	8B-S035-JDG	12 mL (20/box)
	1 g	8B-S035-JEG	20 mL (20/box)
	2 g	8B-S035-KEG	20 mL (20/box)
	5 g	8B-S035-LFF	60 mL (16/box)
96-Well Plate			
	10 mg	8E-S035-AGB	2 Plates/Box
	30 mg	8E-S035-TGB	2 Plates/Box
	60 mg	8E-S035-UGB	2 Plates/Box
96-Well Microelution Plate			
	2 mg	8M-S035-4GA	ea

**Tab-less tubes available. Contact Phenomenex for details.

Luna® PFP(2) HPLC Column Ordering Information

3 µm Minibore Columns (mm)						SecurityGuard™ Cartridges (mm)	
Phases	30 x 2.0	50 x 2.0	100 x 2.0	150 x 2.0		4 x 2.0*	
PFP(2)	00A-4447-B0	00B-4447-B0	00D-4447-B0	00F-4447-B0		AJ0-8326	
						for ID: 2.0-3.0 mm	
3 µm MidBore™ Analytical Columns (mm)						SecurityGuard Cartridges (mm)	
Phases	50 x 3.0	150 x 3.0	50 x 4.6	100 x 4.6	150 x 4.6	4 x 2.0*	4 x 3.0*
PFP(2)	00B-4447-Y0	00F-4447-Y0	00B-4447-E0	00D-4447-E0	00F-4447-E0	AJ0-8326	AJ0-8327
						for ID: 2.0-3.0 mm 3.2-8.0 mm	
5 µm Minibore Columns (mm)						SecurityGuard Cartridges (mm)	
Phases	30 x 2.0	50 x 2.0	150 x 2.0			4 x 2.0*	
PFP(2)	00A-4448-B0	00B-4448-B0	00F-4448-B0			AJ0-8326	
						for ID: 2.0-3.0 mm	
5 µm MidBore™ Analytical Columns (mm)						SecurityGuard Cartridges (mm)	
Phases	50 x 3.0	150 x 3.0	50 x 4.6			4 x 2.0*	4 x 3.0*
PFP(2)	00B-4448-Y0	00F-4448-Y0	00B-4448-E0			AJ0-8326	AJ0-8327
						for ID: 2.0-3.0 mm 3.2-8.0 mm	
5 µm Analytical Columns (mm)						SecurityGuard Cartridges (mm)	
Phases	100 x 4.6	150 x 4.6	250 x 4.6			4 x 3.0*	
PFP(2)	00D-4448-E0	00F-4448-E0	00G-4448-E0			AJ0-8327	
						for ID: 3.2-8.0 mm	

*SecurityGuard™ Analytical Cartridges require holder, Part No.: [KJ0-4282](#)

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Strata-X is patented by Phenomenex. U.S. Patent No. 7,119,145

SecurityGuard is patented by Phenomenex. U.S. Patent No. 6,162,362.

Caution: this patent only applies to the analytical-sized guard cartridge holder, and does not apply to SemiPrep, PREP, or ULTRA holders, or to any cartridges.

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