

# PLICATIONS

### Quantitative Analysis of Gamma Hydroxybutyrate (GHB) in Whole Blood Using Fast SPE and LC-MS/MS

Shahana Huq and Matthew Brusius Phenomenex, Inc., 411 Madrid Ave., Torrance, CA 90501 USA

### Introduction

As a drug of abuse, Gamma-hydroxybutyric Acid (GHB) is often over looked in forensic toxico logical analysis unless it is specifically requested in the screen. Testing for GHB can be difficult due to inaccurate/unreliable response in immunoassays or chromatographic screens and due to the stability of the compound. The time sensitive nature of this assay demands for an analytical procedure that is quick, efficient, and one that preserves the authenticity of the biological specimen. The goal of this application is to present a comprehensive method for the quantitation of GHB in human blood using solid phase extraction (SPE) in conjunction with LC-MS/MS analysis. A quick, two step sample preparation protocol combining protein precipitation (PP) with SPE was employed. The SPE method did not require any conditioning or equilibration of the SPE cartridge, thus allowing for significant time savings. A Luna® 3 µm 150 x 2.0 mm HILIC LC column was utilized for chromatographic analysis, enabling direct injection of the extracted sample in a MS friendly 75% organic. The prescribed protocol circumvents the need for a dry down or reconstitution step and makes it quick and easy to implement in a laboratory workflow, all while providing accurate results.

### Materials and Methods

Reagents and Chemicals

Analytical reference standards and internal standards were purchased from Cerilliant Corporation (Round Rock, TX, USA). Pooled human whole blood with disodium EDTA was purchased from BioreclamationIVT (Westbury, NY). All other reagents and chemicals were purchased from Sigma-Aldrich (St. Louis, MO). Ultrapure D.I water was obtained from Sartorius arium comfort II, courtesy of Sartorious Corporation (Bohemia, NY).

### **SPE Protocol**

Sample Pre-treatment

To 500  $\mu$ L whole blood add 100  $\mu$ L 5 % (w/v) ZnSO<sub>4</sub> and, vortex 3-5 seconds. Add 1.5 mL of chilled (~0 °C) 90:10 Acetonitrile/ Methanol while vortexing. Centrifuge samples at 6000 rpm for 10 minutes and then collect supernatant.

### **SPE Conditions**

96-Well Plate: Strata-X PRO, 30mg/well

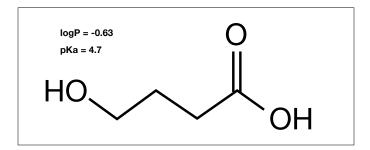
> Part No.: 8F-S536-TGA

> > Pass the supernatant collected from pre-treatment,

apply vacuum and collect extract.

**Direct Injection:** 5 uL of the above sample was injected directly

on LC-MS/MS (bypass dry-down and reconstitution).



### **LC Conditions**

Quantitative Analysis for GHB

Luna® 3 µm HILIC Column: **Dimensions:** 150 x 2.0 mm Part No.: 00F-4449-B0 A: Acetonitrile Mobile Phase:

> B: 100 mM Ammonium Formate Gradient: Time (min) % B 20 20 1.5 50 50 2 01 20

Flow Rate 0.4 mL/min Injection Volume: 5 uL **IHPLC Instrument:** Agilent 1260

MS/MS Instrument: SCIEX® API Triple Quad 4500™, ESI Source (+)

### Qualitative Analysis for Phospholipids

**HPLC Instrument:** 

Kinetex® 2.6 µm C18 Column: Dimensions: 50 x 2.1 mm Part No.: 00B-4462-AN

A: 0.1 % Formic acid in Water Mobile Phase: B: 0.1 % Formic acid in Methanol

**Gradient:** Time (min) % B 40 0.5 95

11.51 40 13.5 Flow Rate 0.4 mL/min Injection Volume: 1 μL

Agilent 1260 MS/MS Instrument: SCIEX API Triple Quad 4500, ESI Source (+)



**Table 1.**Retention time (RT) and MRM Transition for Analytes

			-
Analyte	RT (min)	Q1	Q3
GHB	1.7	104.9	86.9 68.9
GHB-d6	1.7	111	93
Lyso PC	2.4	496.4	184.2
PC-1	6.7	760.7	184.2
PC-2	7.2	786.8	184.2

**Table 2.** % Absolute Recovery of GHB from Extracted Whole Blood Sample (N=4) Using Strata®-X PRO SPE

Spiked Conc.	% Recovery	% CV
10.0 μg/mL	98%	1.7%

**Figure 2.**Representative Chromatogram Showing Phospholipid Trace of Blood Matrix in pre (A) and post (B) SPE (Strata-X PRO) Extracted Sample.

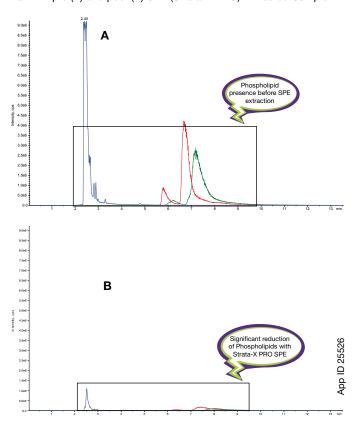
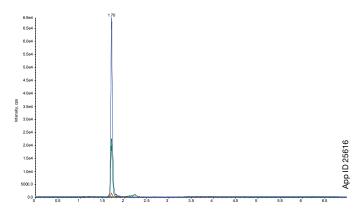


Figure 1.

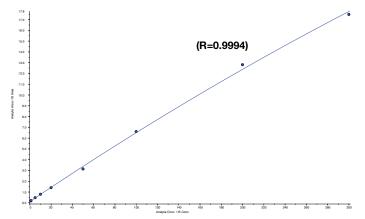
Representative Chromatogram of GHB Extracted Whole Blood Analyzed by a Luna® 3 µm 150 x 2.0 mm HILIC column



**Table 3.** Precision and Accuracy Data for QC Samples

Expected Conc. (μg/mL)	Sample	Replicates (N)	% CV	Accuracy
15	QC 1	4	11.4	104.2
75	QC 2	4	4.5	94.5

**Figure 3.**Linearity Curve for GHB Extracted Blood Matrix over 300-fold Dynamic Concentration Range





### **Results and Discussion**

The developed HILIC LC conditions utilizes a Luna® 3µm HILIC column (Figure 1) and allows for direct injection of extracted samples, bypassing the time consuming dry-down and reconstitution steps, before injection. The method utilizes a protein precipitation followed by a quick sample pass-through SPE method. The resulting extract yields cleaner background (Figure 2B) by selective removal of phospholipids. This is very important because phospholipids can be responsible for unpredictable, inaccurate results in an analytical run as well as causing increased MS instrument down time. The absolute recovery and % CV for extracted blood matrix reported were 98 % and 1.7 % respectively (Table 2). The linear regression value (R=0.9994) of the extracted sample along with precision and accuracy data supports the sound extraction efficiency of the assay over 300-fold dynamic range (Figure 3 and Table 3). The lowest point of linearity curve constructed was 1µg/ mL, as the concentration range around that (< 5ug/mL) point generally is considered endogenous presence rather than GHB ingestion.

## **Ordering Information**

Strata®-X PRO SPE

Format	Sorbent Mass	Part Number	Unit
Tube			
	10 mg	8B-S536-AAK	1 mL (100/box)
unung y	30 mg	8B-S536-TAK	1 mL (100/box)
	30 mg	8B-S536-TBJ	3 mL (50/box)
	60 mg	8B-S536-UBJ	3 mL (50/box)
	200 mg	8B-S536-FBJ	3 mL (50/box)
	100 mg	8B-S536-ECH	6 mL (30/box)
	200 mg	8B-S536-FCH	6 mL (30/box)
	500 mg	8B-S536-HCH	6 mL (30/box)
96-Well Plate			
	10 mg/well	8E-S536-AGA	ea
	30 mg/well	8E-S536-TGA	ea
1=40	60 mg/well	8E-S536-UGA	ea
96-Well Microelution Plat	e		
	2 mg/well	8M-S536-4GA	ea
1000			

### Kinetex® Core-Shell LC Columns

SecurityGuar 2.6 µm Minibore Columns (mm) ULTRA Cartrid							
Phases	30 x 2.1	50 x 2.1	75 x 2.1	100 x 2.1	150 x 2.1	3/pk	
C18	00A-4462-AN	00B-4462-AN	00C-4462-AN	00D-4462-AN	00F-4462-AN	<u>AJ0-8782</u>	
						for 2.1 mm ID	

SecurityGuard 2.6 μm MidBore™ Columns (mm) ULTRA Cartridge								
Phases	30 x 3.0	50 x 3.0	75 x 3.0	100 x 3.0	150 x 3.0	3/pk		
C18	00A-4462-Y0	00B-4462-Y0	00C-4462-Y0	00D-4462-Y0	00F-4462-Y0	<u>AJ0-8775</u>		
						for 3.0 mm ID		

SecurityG 1.7 µm Minibore Columns (mm) ULTRA Cart						
Phases	30 x 2.1	50 x 2.1	100 x 2.1	150 x 2.1	3/pk	
C18	00A-4475-AN	00B-4475-AN	00D-4475-AN	00F-4475-AN	<u>AJ0-8782</u>	
					for 2.1 mm ID	

1.7 µm N	lidBore Column	SecurityGuard ULTRA Cartridges‡	
Phases	50 x 3.0	100 x 3.0	3/pk
C18	00B-4475-Y0	00D-4475-Y0	<u>AJ0-8775</u>
			for 3.0 mm ID

\*SecurityGuard Ultra Cartridges require holder, Part No.: AJO-9000

## Conclusion

The direct injection capability of the Strata®-X PRO extracted sample on the Luna HILIC LC column results in a simple, rapid identification and quantitation of GHB in whole blood. The prescribed method greatly benefits the time sensitive disposition of the assay and clean-up of whole blood matrix to provide an accurate analysis.

### References

- 1. Rachel R. McCusker. Analysis of Gamma-Hydroxybutyrate (GHB) in urine by Gas Chromatography-Mass Spectrometry. J. of Analytical Toxicology. Vol. 23, September 1999
- 2. Po-Chiao Liao a. Clinical management of GHB withdrawal delirium. J. of Formosan Medical Association 117, 1124-1127,
- 3. Fiona J. Couper and Barry K. Logan. Determination of g-Hydroxybutyrate (GHB) in Biological Specimens by Gas Chromatography-Mass Spectrometry. J. of Analytical Toxicology, Vol. 24, January/ February 2000



# ICATIONS

### Luna® LC Columns

3μm Minibore Columns (mm)					yGuard™ Cartridges (mm
Phases	30 x 2.0	50 x 2.0	100 x 2.0	150 x 2.0	4 x 2.0*
					/10pk
HILIC	00A-4449-B0	00B-4449-B0	00D-4449-B0	00F-4449-B0	AJ0-8328
					for ID: 2 0-3 0 mm

3µm MidBor	re™ and Analytical C	SecurityGuard Ca	rtridges (mm)			
Phases	50 x 3.0	150 x 3.0	100 x 4.6	150 x 4.6	4 x 2.0*	4 x 3.0*
					/10pk	/10pk
HILIC	00B-4449-Y0	00F-4449-Y0	00D-4449-E0	00F-4449-E0	AJ0-8328	AJ0-8329
					for ID: 2.0-3.0 mm	3.2-8.0 mm

<sup>\*</sup>SecurityGuard™ Analytical Cartridges require holder, Part No.: KJ0-4282

### Australia

t: +61 (0)2-9428-6444 auinfo@phenomenex.com

**Austria** t: +43 (0)1-319-1301 anfrage@phenomenex.com

**Belgium** t: +32 (0)2 503 4015 (French) t: +32 (0)2 511 8666 (Dutch) beinfo@phenomenex.com

### Canada

t: +1 (800) 543-3681 info@phenomenex.com

### China

t: +86 400-606-8099 cninfo@phenomenex.com

### Denmark

t: +45 4824 8048 nordicinfo@phenomenex.com

### **Finland**

t: +358 (0)9 4789 0063 nordicinfo@phenomenex.com

France t: +33 (0)1 30 09 21 10 franceinfo@phenomenex.com

**Germany** t: +49 (0)6021-58830-0 anfrage@phenomenex.com

t: +91 (0)40-3012 2400 indiainfo@phenomenex.com

### Ireland

t: +353 (0)1 247 5405 eireinfo@phenomenex.com

t: +39 051 6327511 italiainfo@phenomenex.com

Luxembourg t: +31 (0)30-2418700 nlinfo@phenomenex.com

**Mexico** t: 01-800-844-5226 tecnicomx@phenomenex.com

### The Netherlands

t: +31 (0)30-2418700 nlinfo@phenomenex.com

### **New Zealand**

t: +64 (0)9-4780951 nzinfo@phenomenex.com

### Norway

t: +47 810 02 005 nordicinfo@phenomenex.com

**Portugal** t: +351 221 450 488 ptinfo@phenomenex.com

**Singapore** t: +65 800-852-3944 sginfo@phenomenex.com

t: +34 91-413-8613 espinfo@phenomenex.com

### Sweden

t: +46 (0)8 611 6950 nordicinfo@phenomenex.com

### Switzerland

t: +41 (0)61 692 20 20 swissinfo@phenomenex.com

### **United Kingdom**

t: +44 (0)1625-501367 ukinfo@phenomenex.com

t: +1 (310) 212-0555 info@phenomenex.com

## All other countries/regions Corporate Office USA t: +1 (310) 212-0555

info@phenomenex.com

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